

Review Article

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Wide Hybridization in Vegetable Crops

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ABSTRACT

Green revolution has transformed India into a food grain surplus country from a deficit one. No other activity has such an immense impact on the agricultural development as the green revolution has done. It also, has reflected its impact on breeding and production of vegetable crops. Side effects of green revolution were witnessed largely in the form of reduced varietal diversity in major cultivated crop species and increased uniformity in appearance and harvestable products. This predisposed improved agriculture to natural calamities. Emergence of new pathogen races lead to outbreak of diseases and pest attack caused yield losses up to 50 percent. Changing climatic condition has caused abiotic stresses like drought, flood, salinity and high temperature, which in turn results in reduction of yield and quality. To feed the ever-increasing population and to fight with malnutrition, wild species offers the scope for quality improvement in vegetable cultivars.

Keywords

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Introduction

Green revolution has transformed India into a food grain surplus country from a deficit one. No other activity has such an immense impact on the agricultural development as the green revolution has done. It also, has reflected its impact on breeding and production of vegetable crops. Side effects of green revolution were witnessed largely in the form of reduced varietal diversity in major

cultivated crop species and increased uniformity in appearance and harvestable products. This predisposed improved agriculture to natural calamities. Emergence of new pathogen races lead to outbreak of diseases and pest attack caused yield losses up to 50 percent. Changing climatic condition has caused abiotic stresses like drought, flood, salinity and high temperature, which in turn results in reduction of yield and quality. To feed the ever-increasing population and to

fight with malnutrition, wild species offers the scope for quality improvement in vegetable cultivars.

In order to restore the characteristic of ecological sustainability and to combat the biotic and abiotic stress in cultivated vegetable crops, wide hybridization has been advocated a strong tool in the hand of plant breeders as wild species are the rich pool of noble characters, better quality and processing traits as well as imparting resistance against biotic and abiotic stress. Crop wild relatives have been used from decades for breeding, in particular to transfer genes of resistance or tolerance to pests, diseases or abiotic stress to the cultivated species. Wide hybridization comprises the efficient conventional breeding and modern molecular techniques as its effective tool in crop improvement (Table 1–8).

Wide hybridization

Wide hybridization as a norm is an attempt of intermating two species of a genus or two genera of a taxon with an intention of introgression of genes of economic value into the cultivated species. Wide hybridization invariably comprises crosses between wild, primitively cultivated species and genera. Interspecific hybridization means hybridization between individuals from different species belonging to same genus. Intergeneric hybridization means hybridization between individuals from different genus belonging to same family (Liu *et al.*, 2014).

Interspecific and Intergeneric hybridization

Role of wide hybridization

Disease resistance against various pathogens

Insect resistance
Abiotic stress resistance
Quality improvement
Yield enhancement
Development of new crop species
Transfer of sterile cytoplasm for hybrids production
Rootstock breeding

Barriers associated with wide hybridization

Spatial isolation

It is associated with geographical distance, physical separations of time and environment. Sensitivity to photoperiod, introduction to different latitudes and separation by geographical and political barriers can isolate two species reproductively. Development of different maturity groups in knol khol, radish and cauliflower are consequences of spatial isolation.

Pre-fertilization barriers

These barriers are operative between the parental species. They prevent crossability by hindering the fertilization process. Such disturbance of fertilization in interspecific crosses is also termed as interspecific incompatibility or incongruity. Lack of pollen-stigma recognition is mainly responsible for incongruity. The reaction is sporophytic in nature and reported to be associated with pollen wall substances.

Failure of pollen germination

Interspecific hybrid in *Cucumis* species through conventional breeding procedures was unsuccessful because of the existence of a pre-fertilization barrier. The barrier was characterized by non-germination of pollen even up to 72 hours after pollination (Chatterjee and More, 1991). Kaneko and

Bang (2014) reported the same problem in case of *Brassica campestris* and *Raphanus sativus*.

Swollen pollen tube growth

This has been widely reported in case of failure of wide crosses. Incompatibility in *Cucumis spp.* is characterized by delayed growth of pollen, or arrested pollen tube growth in the stigma, or inability of pollen tubes to reach the ovules (Chen and Adelberg, 2000). Based on pollen tube growth behaviour, crosses between African groups of *Cucumis* were classified into three groups, namely bilateral congruity, bilateral incongruity and unilateral incongruity.

Lack of fertilization

In such cases, pollen tube effectively delivered the two sperm nuclei to embryo sac. In one case, zygote was formed but no endosperm development while in second case 10 percent crosses developed endosperm but no embryo.

Post fertilization barriers

These barriers are operative in the distant hybrids and their progenies. These are also known as post syngamic barriers and include embryonic breakdown, failure of zygote development, abnormal fertilization, inhibition of endosperm and embryo development.

Hybrid embryo abortion

Arrest of embryo development or its abortion has been noticed in several interspecific hybrids. The major barriers to interspecific hybridization in *Phaseolus sp. i.e. Phaseolus vulgaris* × *P. acutifolius* is embryo abortion. In case of *Phaseolus coccineus* × *P. vulgaris*, abnormal embryo development is observed (Andradf- Aguilar and Jackson, 1988).

Hybrid inviability

It results from various factors and manifests from zygote development to seed formation. It operates in F₁ seedlings causing high mortality, hybrid plants are characterized by poor growth, chlorotic leaves and die before maturity. The reason may be disharmony between nucleus of one species and cytoplasm of other or between nucleus of both the species or due to action of specific genes known to cause lethality, chlorosis and weakness in F₁ hybrids. e.g. Cross between *Cucumis sativus* × *C. melo* and *Abelmoschus esculentus* × *A. tetraphyllus*.

Hybrid sterility

F₁ hybrids are characterized by lack of seed set. Hybrid sterility is due to manifestation of lethal genes, genetic imbalance due to chromosomal non-homology, chromosomal elimination and endosperm abortion. Several interspecific hybrids of *Abelmoschus esculentus* × *A. ficulneus* and *Abelmoschus esculentus* × *A. tuberculatus* show very poor seed set.

Failure of flowering in the progenies

Either hybrid is devoid of reproductive structure or highly deformed non-functional structures are formed. It is due to failure of physiological differentiation while non-functional reproductive systems are attributed to failure of meiosis in either one or both the sexes.

Hybrid breakdown

It is manifested in the form of inviable and weak F₂ generation or later generation. In some wide crosses seed setting is normal, even F₁ plant progenies show normal development and good fertility but in the F₂ generation performance of hybrids fall below satisfaction and sometimes may be complete

sterility. Afful *et al.*, (2018) observed failure in fruit set, when the wild accessions were used as female parents in the crossability study of cultivated brinjal with *Solanum torvum*, *S. anguvi* and *S. aethiopicum*.

Techniques to overcome pre-fertilization barriers

Taxonomic position of parental species

An ease in hybridization is expected when the species more resemble phenotypically. Taxonomic classification is based on morphological features but by and large is the outcome of genetic factors in association with environment.

Doubling of chromosome number

When the failure of hybridization is due to different ploidy level then this technique is followed. In many polyploid cultivated species, their wild progenitors are diploid and crossing attempts are difficult. One may increase ploidy level of wild type by colchicine treatment. This enhances the success rate in crops like potato, *Cucumis* and *Brassica* spp.

Bridging species technique

In many crops, two species which are otherwise incompatible, may be hybridized with the help of third species. The third species acts as bridge in recombining the two incompatible species so known as bridging species. This technique has been used in making wide crosses in potato, lettuce and sugar beet.

Hayes *et al.*, (2005) were successful in using *S. verrucosum* as a female bridging parent to access 2x (1EBN) *S. pinnatisectum*. *Solanum simplicifolium* was used as bridging species in crossing *S. acuale* and *S. tuberosum*

Shortening of the style

In some species, the incompatible reaction can be overcome by reducing the length of style. Incompatible reaction in radish can also be overcome by reduction of stylar tissues.

Mentor pollination

Pollen grains of distant species do not germinate on the stigma of cultivated species. However, when these pollen grains are mixed with killed maternal pollen grains, germination of the incompatible pollen grains take place as in case of *Cucumis* (Beharav and Cohen, 1994). It happens because cell wall proteins of pollen play pivotal role in pollen-pistil interaction. Pollen killed in ethanol and mixed with incompatible pollens, release the proteinaceous recognition factors, thereby masking rejection reaction of the recipient stigma. The killed maternal pollen is known as mentor pollen.

Use of growth regulators and immunosuppressants

Growth Regulators (GRs) enhance the zygotic formation in distant hybrids of many cucurbits, okra and tomato. Commonly used GRs are IAA, IBA, GA₃. GA₃ 75ppm application to maternal plant 1 or 2 days before and after pollination improved zygote formation, faster pollen tube growth, more embryo survival and more seed set in wide crosses. Application of auxin in *Solanum* prevented flower abscission and enhanced fruit set.

In vitro fertilization

It is effective when stigma and style inhibit pollen tube growth and embryo abortion occurs at early stages of development. The whole gynoecium is excised and placed on MS medium followed by dusting of pollens on the stigma and fertilized ovule is reared to maturity (Ondrej *et al.*, 2002).

Protoplast fusion

It involves the fusion of protoplast of two incompatible species or genera and the fused heterokaryon is placed on artificial nutrient medium and regenerated into hybrid plant. This approach has particularly more potential to generate new genotypes in vegetatively propagating species. Some examples are tomato+potato, *Solanum nigrum*+ *S. tuberosum*. Chandel *et al.*, 2015 studied interspecific potato somatic hybrids between cultivated *S. tuberosum* dihaploid C-13 and wild species *S. cardiophyllum* via protoplast fusion. The use of somatic fusion for improving valuable agronomic traits in cultivated potatoes is done for traits like atrazine resistance, frost tolerance, quality improvement and pest and diseases resistance.

Techniques to overcome post-fertilization barriers

Embryo rescue

Embryo abortion is the major barrier in distant hybridization. It can occur at any stage of development depending on the genomic relationship of two parental species. *Solanum sitiens* is a rare endemic plant of the Atacama Desert of Chile having tolerance to drought, salinity and low temperatures, resistances to certain pathogens, and modified fruit ripening. To overcome sterility and unilateral incompatibility of *Solanum lycopersicum* × *S. sitiens* hybrids, embryo culture was used (Chetelat, 2016).

Capsicum chinense, *C. annuum* and *C. frutescens* were crossed with each other and embryo rescue was done between 27-33 days after pollination by Debbarama *et al.*, 2013. Hybrid plants were obtained and their hybridity was confirmed using both morphological and RAPD markers.

Ovary culture

In some wide crosses, embryo abortion occurs at early stages of development when it is difficult to excise and culture embryos. To overcome this problem, ovaries are cultured. Depending on cross combination, ovaries 2-15 days after pollination are excised and cultured on nutrient medium. Flower is pruned by removing calyx, corolla and stamen. Distal part of the pedicel is cut and ovary planted on simple semi-solid MS nutrient medium. When the embryos become visible, they are aseptically taken out and cultured in a manner of embryo rescue technique

Ovule culture

Ovule culture is an elegant experimental system by which ovules are aseptically isolated from the ovary and are grown aseptically on chemically defined nutrient medium under controlled conditions. It is used when barrier impedes growth of zygote at earlier stages of development.

Backcrossing

Wide crosses showing poor fertility can be back crossed to cultivated species to improve the fertility of hybrids. Backcrossing can be applied to balance cytoplasmic interaction by producing cybrids or alloplasmic lines.

Chamola *et al.*, 2015 used cytoplasmic male sterile (CMS) lines of *Brassica juncea* and *B. napus* with the mitochondrial genome of *Moricandia arvensis* and *Erucastrum canariense*, respectively, were used to transfer CMS to cauliflower (*B. oleracea*). Embryo rescue was also done in BC1 and BC2 to obtain progenies. Recovery of the recurrent parent phenotype was faster in *B. napus* × *B. oleracea* than *B. juncea* × *B. oleracea*. BC3 generation plants of *B. napus* × *B. oleracea*

showed good curd formation and complete male sterility and nine bivalents at meiosis whereas those of *B. juncea* x *B. oleracea* were male sterile but still had genetic elements of *B. juncea*.

Table.1 Interspecific v/s Intergeneric hybridization

Particulars	Interspecific Hybridization	Intergeneric hybridization
Parents involved	Involve two different species of the same genus	Involve two different genera of the same family
Fertility	Such hybrids vary from completely fertile to completely sterile	Such hybrids always sterile
Use in crop improvement	Frequently used	less than interspecific crosses
Release of hybrid Varieties	Possible in some crops	Not possible
Evolution of new crops	Not possible, but evolution of new species is sometimes possible	Sometimes possible

Table.2 Disease resistance in different vegetable crops

Crop	Character transferred	Wild Species	Species
Okra	Resistance to YVMV	<i>Abelmoschus caillei</i>	<i>A. esculenta</i>
Brinjal	Bacterial wilt	<i>S. stenotomum</i>	<i>S. melongena</i>
Tomato	Fusarium wilt	<i>Solanum hirsutum</i>	<i>S. lycopersicum</i>
Chilli	Fruit rot	<i>Capsicum chinense</i>	<i>C. annum</i>
Onion	Purple blotch	<i>Allium fistulosum</i>	<i>A. cepa</i>
Potato	Late blight, leaf roll, virus-x	<i>Solanum demissum</i>	<i>S. tuberosum</i>
French bean	Rust resistant	<i>P. flavescens</i>	<i>P. vulgaris</i>
Cucumber	Green-mottle mosaic	<i>Cucumis hardwickii</i>	<i>C. sativus</i>

Table.3 Insect resistance in different vegetable crops

Crop	Character transferred	Species transferred from	Species transferred to
Tomato	White fly	<i>Solanum hirsutum</i>	<i>S. esculentum</i>
	Root knot nematode	<i>S. peruvianum</i>	<i>S. esculentum</i>
Potato	Nematode	<i>S. vernei</i>	<i>S. tuberosum</i>
Okra	Fruit and shoot borer	<i>A. manihot</i>	<i>A. esculentus</i>
Brinjal	Shoot and fruit borer	<i>S. incanum</i>	<i>S. melongena</i>
	Epilachna beetle	<i>S. nigrum</i>	<i>S. melongena</i>
Cucurbits	Fruit fly	<i>Cucumis trigonus</i>	<i>C. sativus</i>

Table.4 Abiotic stress resistance in different vegetable crops

Crop	Character transferred	Species transferred from	Species transferred to
Tomato	High temperature	<i>Solanum cheesmani</i>	<i>S. lycopersicum</i>
Potato	Frost tolerance	<i>Solanum acaule</i>	<i>S. tuberosum</i>
	Heat and drought	<i>Solanum chacoense</i>	<i>S. tuberosum</i>
Onion	Tolerance to cold	<i>Allium porrum</i>	<i>A. cepa</i>
Cucumber	Salinity	<i>Benincasa hispida</i>	<i>Cucumis sativus</i>
Brassicae	Drought and heat	<i>Brassica chinensis</i>	<i>B. oleraceae</i>
Okra	Low temperature	<i>Abelmoschus angulosus</i>	<i>A. esculentus</i>

Table.5 Quality improvement in different vegetable crops

Crop	Character transferred	Species transferred from	Species transferred to
Tomato	Carotenoid content	<i>Solanum hirsutum</i>	<i>S. esculentum</i>
	Soluble solid	<i>Solanum chmielewskii</i>	<i>S. esculentum</i>
Melon	Thick rind and good keeping quality	<i>Cucumismelovar. cantaloupensis</i>	<i>C. melo</i>
Potato	Starch content	<i>Solanum acaule</i>	<i>S. tuberosum</i>
Chilli	High capsaicin	<i>C. frutescence</i>	<i>C. annum</i>
Onion	Leaf flavour	<i>Allium kurrat</i>	<i>A. cepa</i>

Table.6 The use of somatic fusion for improving valuable agronomic traits in cultivated potatoes

Trait	Fusion partner
Atrazine resistance	<i>S. tuberosum</i> (+) <i>S. nigrum</i>
Frost tolerance	<i>S. tuberosum</i> (+) <i>S. commersonii</i>
Reduced glycoalkaloid aglycones	<i>S. tuberosum</i> (+) <i>S. brevidens</i>
Resistance to bacterial wilt	<i>S. tuberosum</i> (+) <i>S. phureja</i>
Resistance to late blight	<i>S. tuberosum</i> (+) <i>S. brevidens</i> ; <i>S. tuberosum</i> (+) <i>S. bulbocastanum</i>
Resistance to potato leaf roll virus (PLRV)	<i>S. tuberosum</i> (+) <i>S. brevidens</i> ; <i>S. tuberosum</i> (+) <i>S. verrucosum</i>
Resistance to potato virus Y (PVY)	<i>S. tuberosum</i> (+) <i>S. brevidens</i>

Table.7 Inter-specific hybridization in vegetable crops

Tomato	Pusa Red Plum	<i>S. lycopersicum</i> x <i>S. pimpinellifolium</i>	Rich in Vit-C
	Hissar Anmol	<i>Hissar Arun</i> x <i>S. hirsutum</i> f. <i>glabratum</i>	Resistant to TLCV
Potato	Kufri Kuber	(<i>Solanum curtilobum</i> x <i>S. tuberosum</i>) x <i>S. andigenum</i>	High tuber yield
Cucumber	C. hytivus	<i>Cucumis sativus</i> x <i>C. hystrix</i>	Resistant to downy mildew
Amaranthus	Pusa Kiran	<i>A. tricolour</i> x <i>A. tristis</i>	Rainy season
Okra	Pusa A4	<i>A. esculentus</i> x <i>A. manihot</i> ssp. <i>manihot</i>	Tolerant to jassids, Fruit and shoot borer
	Punjab-7	<i>A. esculentus</i> (Pusa Sawani) x <i>A. manihot</i> ssp. <i>manihot</i> (Ghana)(2n=194)	Resistance to YVMV
	Punjab Padmini	<i>A. esculentus</i> (Rashmi) x <i>A. manihot</i> ssp. <i>manihot</i> (Ghana)	
	Parbhani Kranti	<i>A. esculentus</i> (Pusa Sawani) x <i>A. manihot</i>	
	Arka Anamika	<i>A. esculentus</i> (2n=130) x <i>A. tetraphyllus</i> (2n=138)	
	Arka Abhay	<i>A. esculentus</i> (2n=72) x <i>A. tetraphyllus</i> (2n=130)	Resistance to YVMV and tolerant to fruit borer

Table.8 Inter-generic hybridization in vegetable crops

New Crop	Parents	Special feature
Hakurana	Cabbage x Chinese cabbage (Developed by Embryo culture)	New leafy vegetable in Japan Resistant to soft rot, drought and heat
Nabicol	Kale x Turnip	
Caulicob	Cabbagex Cauliflower	
Swede	Turnip x Cabbage	Root vegetable
<i>Raphanobrassica</i>	Radish x Cabbage	Fodder crop
Baemoochae (<i>Brassicoraphanus</i>) (2n=38)	<i>B. rapa</i> ssp. <i>pekinensis</i> (Big head Chinese cabbage) x <i>R. sativus</i> (big root radish)	New leafy vegetable

Chromosome doubling

Distant hybrids exhibit high degree of sterility, which is mainly due to genic or chromosomal difference between parental species. This problem in many cases has been overcome by doubling the chromosome number of F₁ hybrid. Bharathi *et al.*, (2014)

crossed natural tetraploid *Momordica subangulata* subsp. *renigera* (2n = 56) with induced tetraploid *Momordica dioica* (2n = 4x = 56) to produce new species *Momordica x suboica* Bharathi. This hybrid maintains its morphological characteristics, superior agronomic traits making it a good choice as a new vegetable crop.

In conclusion, wild species are the store house of novel genes, which can be utilized for wide hybridization. So, wild species forms the heart of wide hybridization programme. Wide hybridization is an effective tool for genetic improvement in vegetables crops. It is a highly technical and knowledge intensive process. Even though, wide hybridization has so many barriers, it can be overcome through the conventional and biotechnological approach. To broaden the genetic base, wide hybridization following *in vitro* and biotechnological approach is being used. It can be used for disease and insect resistance breeding and quality improvement in the cultivars, which is need of the hour.

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